

Sunniest C18, C18-HT, RP-AQUA, C8, PhE, PFP

A Novel Bonding Technique

(patent pending)

An unique trifunctional silyl-reagent was developed as shown in Fig. 1. This silyl-reagent can bond with any silanol groups on silica surface as shown in Fig.2 because it can expand and contract by itself. This technique can make residual silanol groups on C18 stationary phase to be the least amount.

Finally an end-capping was done with trimethylsilyl-reagent (TMS).

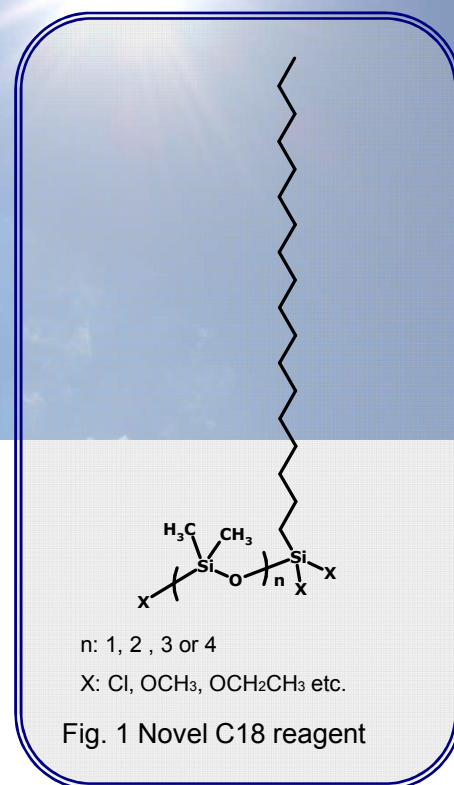


Fig. 1 Novel C18 reagent

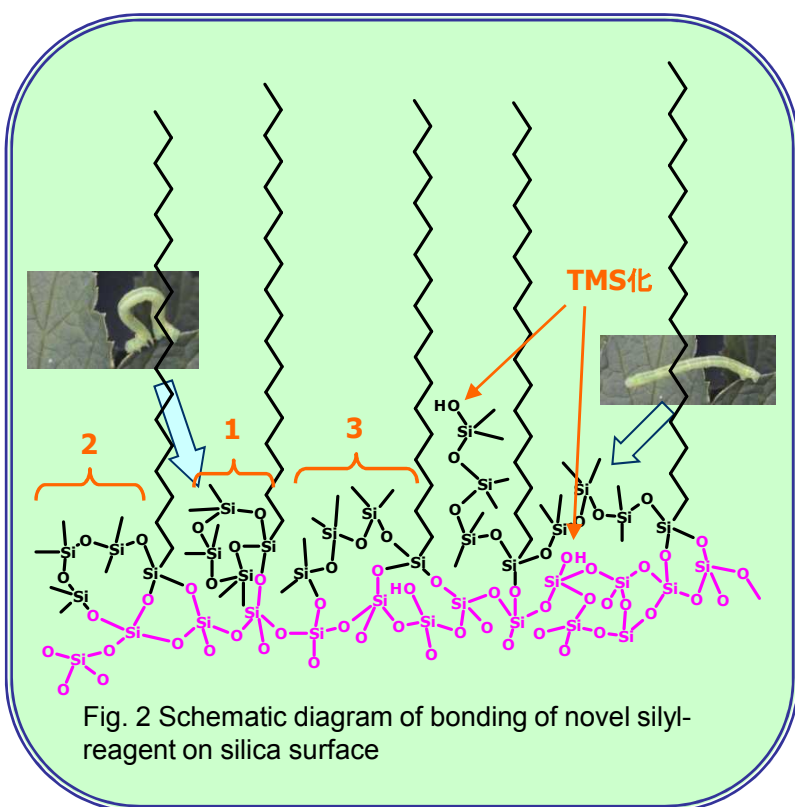


Fig. 2 Schematic diagram of bonding of novel silyl-reagent on silica surface

Features

- ★ Little residual silanol groups by an unique bonding technique
- ★ Excellent stability, especially under acidic pH conditions
- ★ Sharp peak shape for acidic, basic and chelating compounds
- ★ RP-AQUA is available under 100% aqueous conditions, and shows enhanced retention of polar compounds.

Characteristics of Sunniest

| | Particle size (μm) | Pore diameter (nm) | Specific surface area (m ² /g) | Carbon content (%) | Bonded phase | pH range |
|------------------|--------------------|--------------------|---|--------------------|-------------------|----------|
| Sunniest C18 | 3 and 5 | 12 | 340 | 16 | C18 | 1.5 - 10 |
| Sunniest C18-HT | 2 | 10 | 340 | 16 | C18 | 1.5 - 10 |
| Sunniest RP-AQUA | 3 and 5 | 12 | 340 | 16 | C28 | 2 - 8 |
| Sunniest C8 | 3 and 5 | 12 | 340 | 10 | C8 | 1.5 - 9 |
| Sunniest PhE | 3 and 5 | 12 | 340 | 10 | Phenylethyl | 1.5 - 8 |
| Sunniest PFP | 5 | 12 | 340 | 10 | Pentafluorophenyl | 2 - 8 |

Sunniest C18

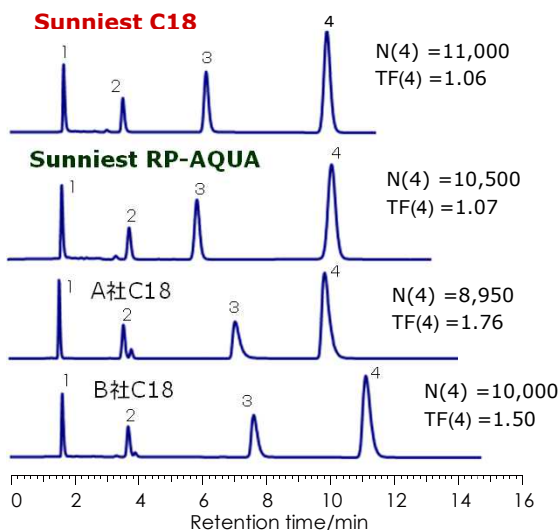
Sunniest RP-AQUA

Sunniest C8

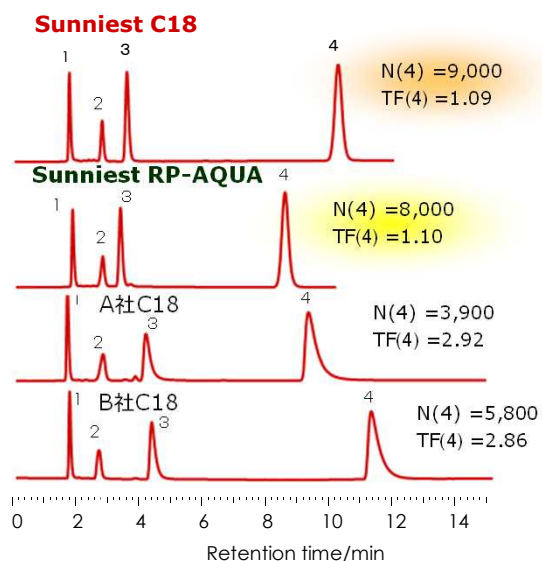
◆Evaluation of End-capping

Comparison of plates number (N) and USP tailing factor (TF) of amitriptyline

メタノール CH₃OH, pH7.5, 40 °C



メタノール CH₃OH, pH6.0, 22 °C



Column size: 4.6 x 150 mm

Particle size: 5 μm

Mobile phase:

CH₃OH/20mM Phosphate buffer pH7.5 or 6.0 =80/20

CH₃CN/20mM Phosphate buffer pH7.0 =60/40

Flow rate: 1.0 mL/min

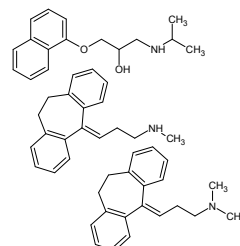
Temperature: 40 °C or 22 °C

Sample: 1 = Uracil

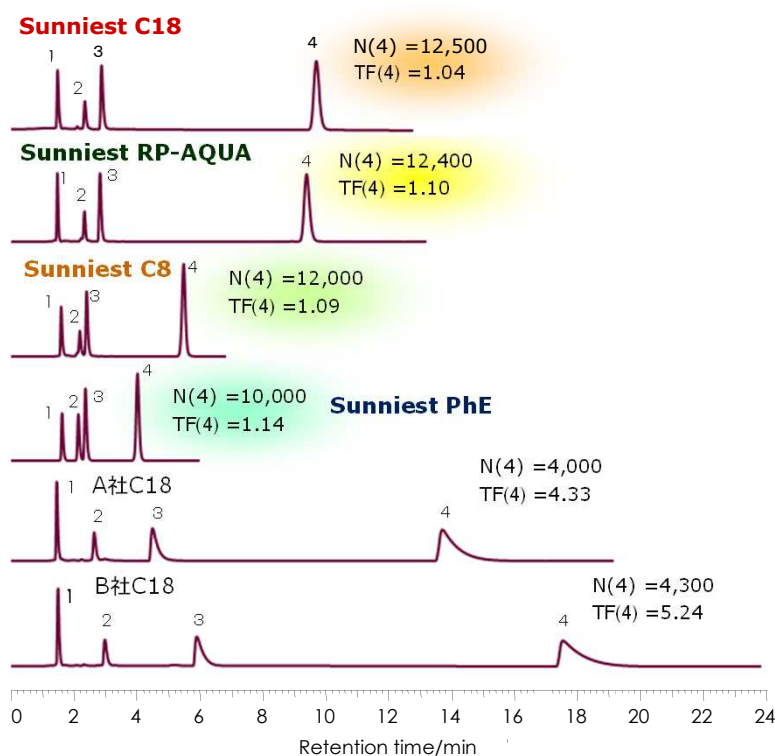
2 = Propranolol

3 = Nortriptyline

4 = Amitriptyline



アセトニトリル CH₃CN, pH7.0, 40 °C



Amitriptyline is widely used to evaluate residual silanol groups on the C18 stationary phase. Peak shape of amitriptyline was compared under 3 kinds of conditions such as methanol/phosphate buffer/40 °C, methanol/phosphate buffer/22 °C and acetonitrile/phosphate buffer/40 °C. Under methanol/phosphate buffer/40 °C conditions which is the most common among HPLC manufacturers, good peak shape was obtained for all columns. There were little difference of a peak shape. Under acetonitrile/phosphate buffer/40 °C, however, Sunniest columns showed a symmetrical peak, while column A and B C18 showed a terribly tailing peak.

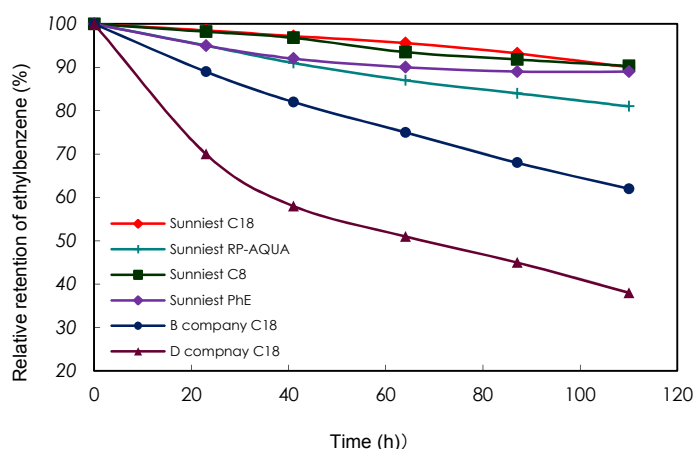
Sunniest C18, RP-AQUA and C8 columns allow to use a wide range of mobile phase without peak tailing because of extremely low content of residual silanol groups on the stationary phase.

Sunniest C18

Sunniest RP-AQUA

Sunniest C8

◆ Stability under acidic and basic pH conditions

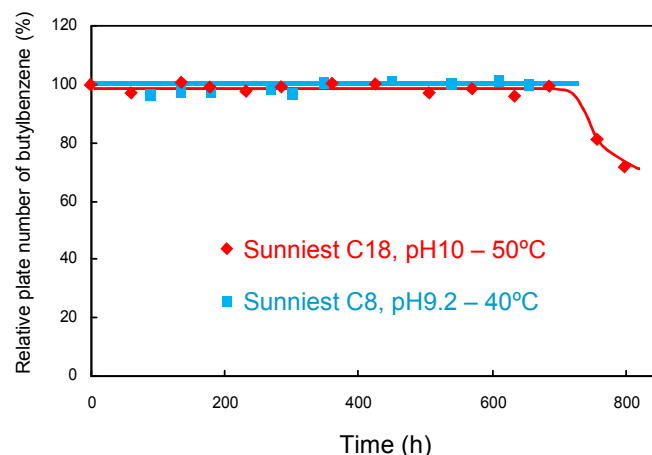


Durable test condition

Column size: 150 x 4.6 mm
 Mobile phase: CH₃CN/1.0% TFA (pH1) = 10/90
 Flow rate: 1.0 mL/min
 Temperature: 80 °C

Measurement condition

Column size: 150 x 4.6 mm
 Mobile phase: CH₃CN/H₂O=60/40
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: 1 = Uracil
 2 = Ethylbenzene



Durable test condition

Column: Sunniest C18, C8, 5 μm 150 x 4.6 mm
 Mobile phase:
 C18: CH₃OH/20mM Sodium borate/10mM NaOH=30/21/49 (pH10)
 C8: CH₃OH/20mM Sodium borate (pH9.2) =30/70
 Flow rate: 1.0 mL/min
 Temperature: C18 - 50 °C, C8 - 40 °C

Measurement condition

Column: Sunniest C18, C8, 5 μm 150 x 4.6 mm
 Mobile phase: CH₃OH/H₂O=75/25
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: 1 = Butylbenzene

Stability under acidic pH conditions was evaluated at 80 °C using acetonitrile/1% trifluoroacetic acid solution (10:90) as mobile phase. 100% aqueous mobile phase expels from the pore of packing materials by capillarity and packing materials doesn't deteriorate. 10% acetonitrile in a mobile phase allows an accurate evaluation.¹⁻³⁾

★ Sunniest C18 has kept 90% retention for 100 hours under such severe conditions.

Our bonding technique can make column life be long.

Sunniest RP-AQUA is less stable than Sunniest C18. However, Sunniest RP-AQUA has more stable than other company C18 columns.

Stability under basic pH conditions was evaluated at 50 °C using methanol/Sodium borate buffer pH 10 (30:70) as mobile phase. Sodium borate is used as an alkaline standard solution for pH meter, so that its buffer capacity is high.

Elevated temperature of 10 °C makes column life be one third. When Sunniest C18 column is used at 40 °C, column life becomes 2,000 hours. Other company shows stability test at ambient (room temperature). If room temperature is 25 °C, column life at room temperature (25 °C) is sixteen times longer than that at 50 °C.

★ Sunniest C18 is enough stable even if it is used under pH 10 condition. Regarding stability under basic pH condition, there is little C18 column like Sunniest C18 except for hybrid type C18. It is considered that our end-capping technique leads high stability.

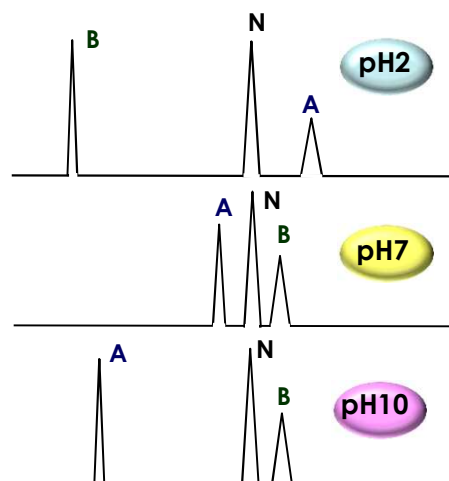
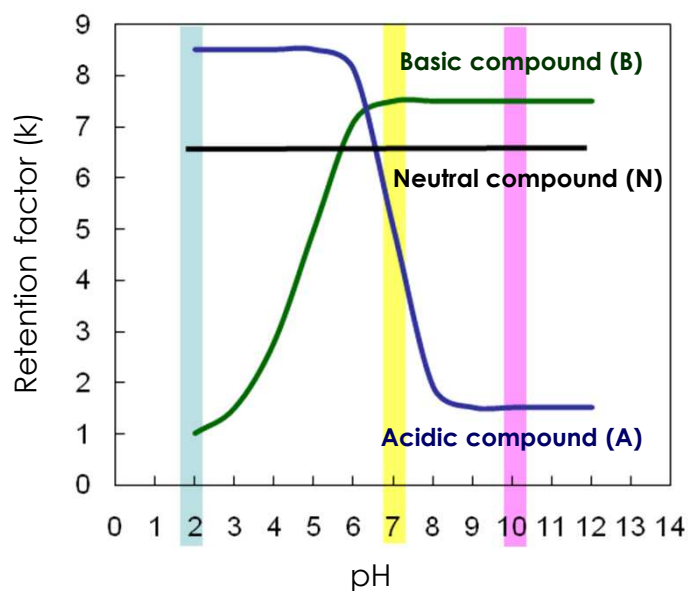
★ Sunniest C18 can be used at the pH range from 1.5 to 10. Sunniest C8 can be used at the pH range from 1.5 to 9.

1) N. Nagae, T. Enami and S. Doshi, LC/GC North America October 2002.

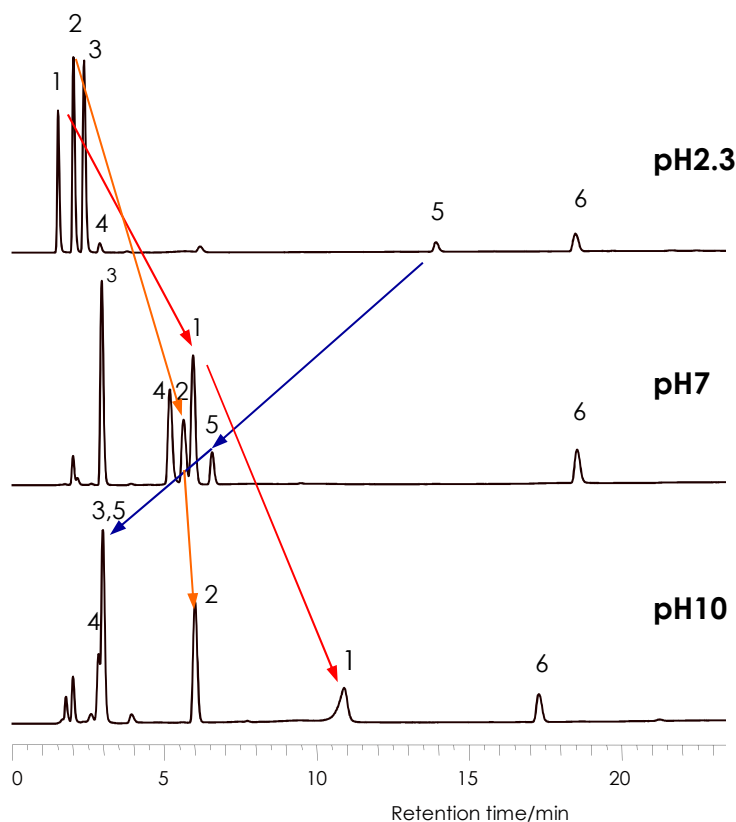
2) T. Enami and N. Nagae, American Laboratory October 2004.

3) T. Enami and N. Nagae, BUNSEKI KAGAKU, 53 (2004) 1309.

◆ Relationship between pH and retention of acidic, basic and neutral compounds



◆ pH selectivity



Column: Sunniest C18, 5 μ m 150 x 4.6 mm

Mobile phase:

A1) 20mM Phosphoric acid pH2.3

A2) 20mM Phosphate buffer pH7

A3) 20mM Phosphate buffer pH10

B) Acetonitrile

Time (min) 0 30

%B (%) 2 26

Flow rate: 1.0 mL/min

Temperature: 40 $^{\circ}$ C

Detection: UV@250 nm

Sample: 1 = Thiamine HCl Vitamin B₁

2 = Nicotinamide

3 = Nicotinic acid

4 = Pyridoxine HCl Vitamin B₆

5 = Folic acid

6 = Riboflavin Vitamin B₂

pH of mobile phase can make selectivity of ionic compounds change much. Sunniest C18 can be used at the pH range from 1.5 to 10, so that a suitable analytical method can be created using Sunniest C18.

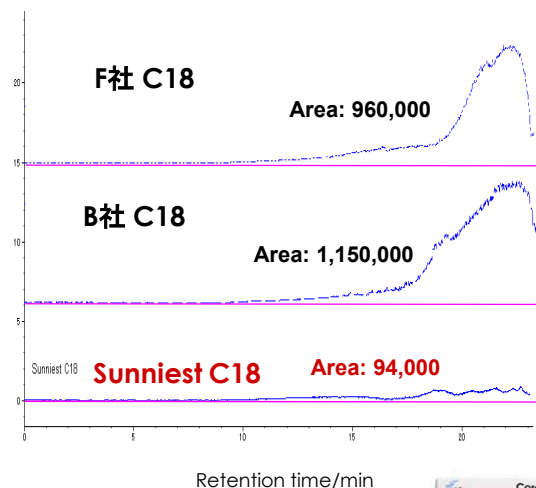
Sunniest C18

Sunniest RP-AQUA

Sunniest C8

◆Comparison data: Bleeding from column

《Comparison using Corona CAD》



Column size: 150 x 2.0 mm

Mobile phase:

A) 0.1% acetic acid

B) CH₃CN

Gradient:

Time: 0min 3min 14.4min 18min 19min

%B: 5% 5% 100% 100% 5%

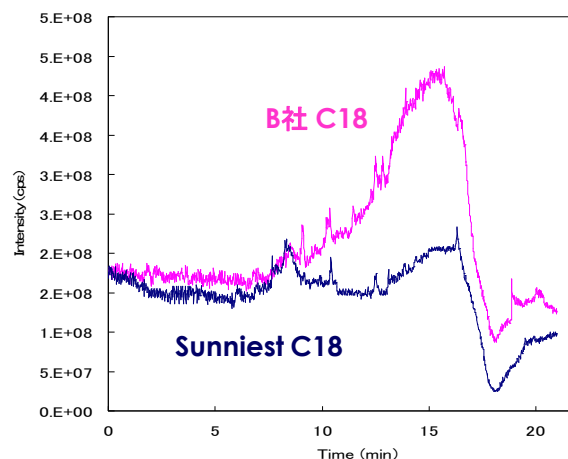
Flow rate: 0.2 mL/min

Temperature: 40 °C

Detection: Corona CAD



《Comparison using MS》



Column size: 150 x 2.0 mm

Mobile phase:

A) 0.1% acetic acid

B) CH₃CN

Gradient: Time: 0min 3min 14.4min 18min 19min

%B: 5% 5% 100% 100% 5%

Flow rate: 0.2 mL/min

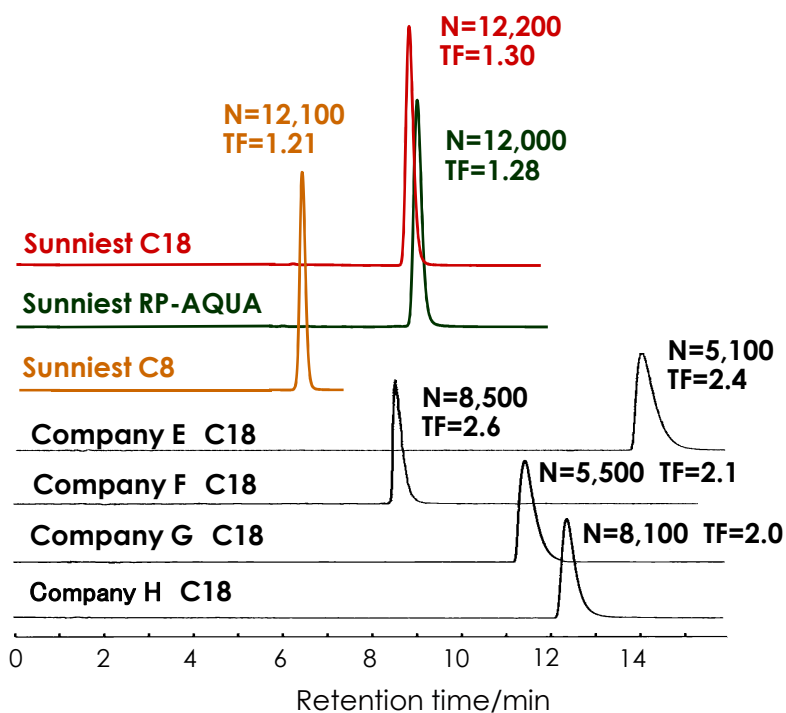
Temperature: 40 °C

MS: ABI API-4000

Ionization: Turboionspray (cation)

Measurement mode: Q1 Scan m/z 100-1000

◆Separation of antidepressants using acetonitrile and ammonium acetate for LC/MS



Column size: 150 x 4.6 mm

Particle size: 5μm

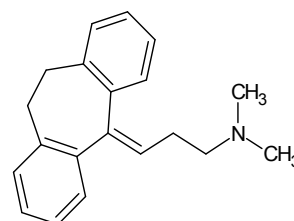
Mobile phase: CH₃CN/10mM

Ammonium acetate pH6.8=40/60

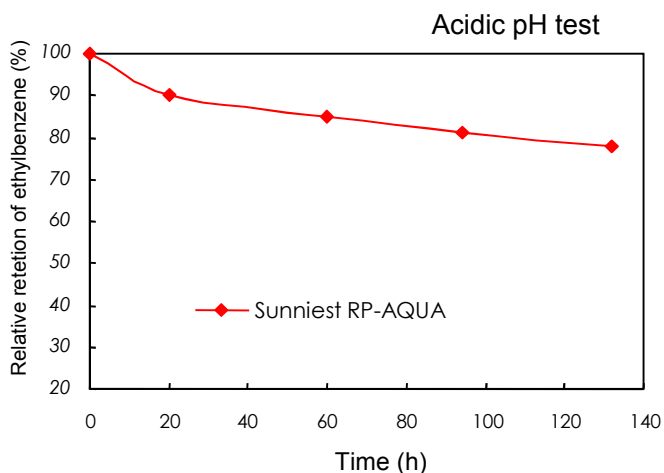
Flow rate: 1.0 mL/min

Temperature: 40 °C

Sample: Amitriptyline



◆ **Stability of Sunniest RP-AQUA under 100% aqueous conditions**

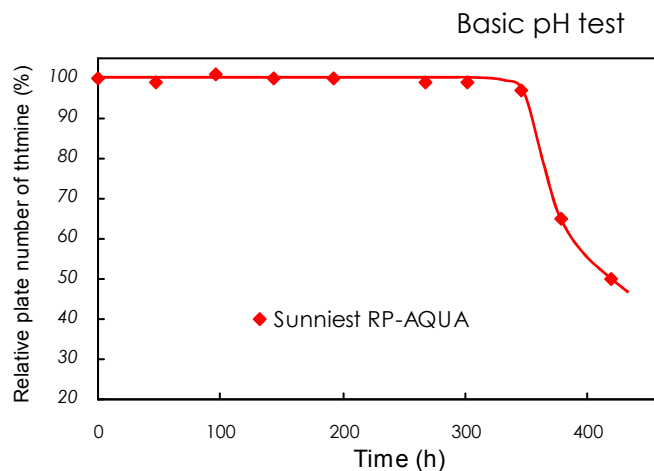


Durable test conditions

Column: Sunniest RP-AQUA, 5 μ m 150 x 4.6 mm
 Mobile phase: 0.5% TFA
 Flow rate: 1.0 mL/min
 Temperature: 60 °C

Measurement conditions

Column: Sunniest RP-AQUA, 5 μ m 150 x 4.6 mm
 Mobile phase:
 CH₃OH/H₂O=75/25
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: 1 = Uracil
 2 = Amylbenzene



Durable test conditions

Column: Sunniest RP-AQUA, 5 μ m 150 x 4.6 mm
 Mobile phase: 20mM Phosphate buffer pH8.0
 Flow rate: 1.0 mL/min
 Temperature: 40 °C

Measurement conditions

Column: Sunniest RP-AQUA, 5 μ m 150 x 4.6 mm
 Mobile phase: 10mM Phosphate buffer pH7.0
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Sample: 1 = Thymine

It is important that stability is evaluated under 100% aqueous conditions for an AQUA type C18 column because column life becomes longer to increase a content of organic solvent in a mobile phase. Sunniest RP-AQUA can be used under 100% aqueous conditions from pH2 to pH8.

★ Sunniest RP-AQUA can be used under 100% aqueous conditions from pH 2 to pH 8. Sunniest RP-AQUA is one of the most stable aqua type column.



◆ Reproducibility of retention under 100% aqueous conditions

★ C18 and C8 reversed phases exhibit decreased and poorly reproducible retention under more than 98% aqueous conditions as shown in Fig. 1. This problem traditionally has been explained as being the result of ligand collapse or a matting effect. Nagae¹⁻³ ascertained, however, that the mobile phase was being expelled from the pores of the packing material under 100% aqueous mobile phase conditions, as Fig. 2 shows.

★ When the surface of packing materials isn't wet by water, water used as a mobile phase expels from the pore of the packing material by capillarity. This is a reason why reproducibility in retention is low under 100% aqueous conditions. Reversely pressure around the packing material makes water permeate into the pore of the packing material to overcome a force worked by capillarity.

In other words, the surface of a reversed phase like C18 isn't wet by water anytime even if water permeates into the pore of the packing material or not. So it is wrong that we say "dewetting" when water expel from the pore. Saying "Depermetating" is more suitable.

★ Sunniest RP-AQUA is a reversed stationary phase, so that it is not wet with water. However the contact angle of water on the surface of Sunniest RP-AQUA is less than that of a conventional C18. Expelling force (pressure) acted by capillarity on Sunniest RP-AQUA is less than atmospheric pressure. So, Sunniest RP-AQUA shows reproducible retention under 100% aqueous conditions.

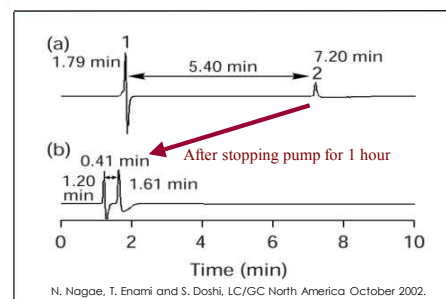


Fig. 1 Retention behavior of a C18 column under 100% aqueous mobile phase conditions

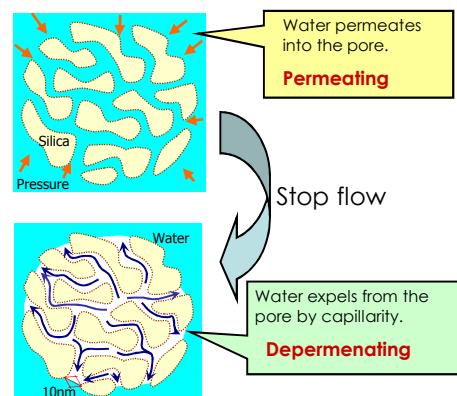
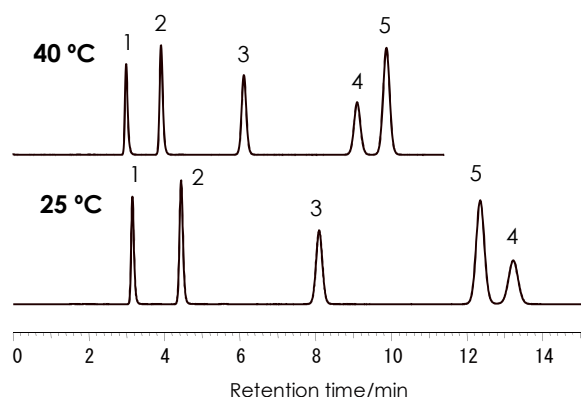


Fig. 2 Schematic diagram of C18 particle

1) N. Nagae, T. Enami and S. Doshi, LC/GC North America October 2002.
3) T. Enami and N. Nagae, BUNSEKI KAGAKU, 53 (2004) 1309.

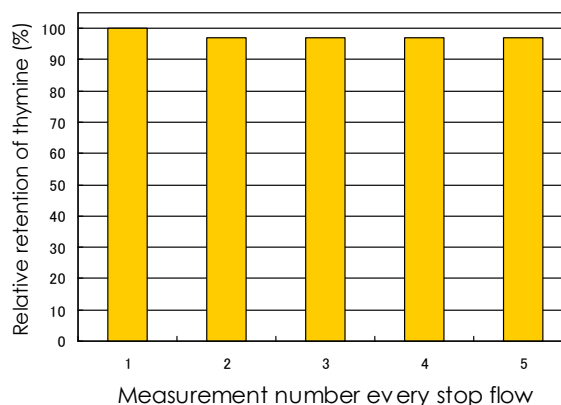
2) T. Enami and N. Nagae, American Laboratory October 2004.

◆ Separation of nucleic acid bases



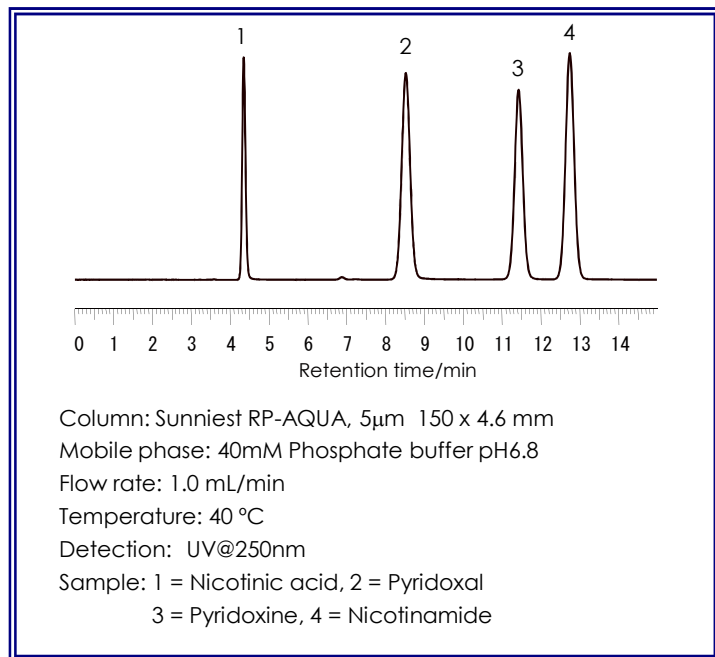
Column: Sunniest RP-AQUA, 5 μ m 150 x 4.6 mm
Mobile phase: 10mM Phosphate buffer pH7.0
Flow rate: 1.0 mL/min
Temperature: 40 °C and 25 °C
Sample: 1 = Cytosine 2 = Uracil
3 = Thymidine 4 = Uridine
5 = Thymine

Change of retention of thymine at 40 °C (measurement every stop flow for 1 hour)

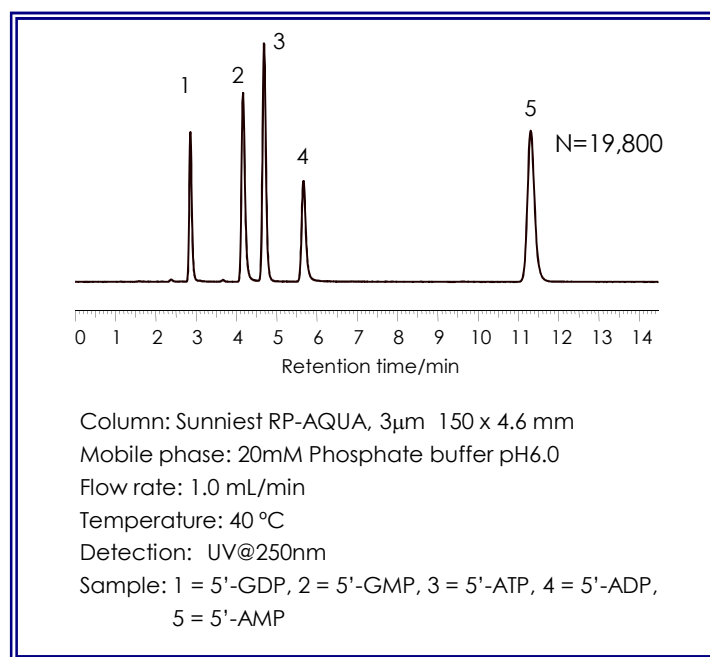


Sunniest RP-AQUA showed more than 97% of reproducibility in retention using 100% aqueous buffer as a mobile phase.

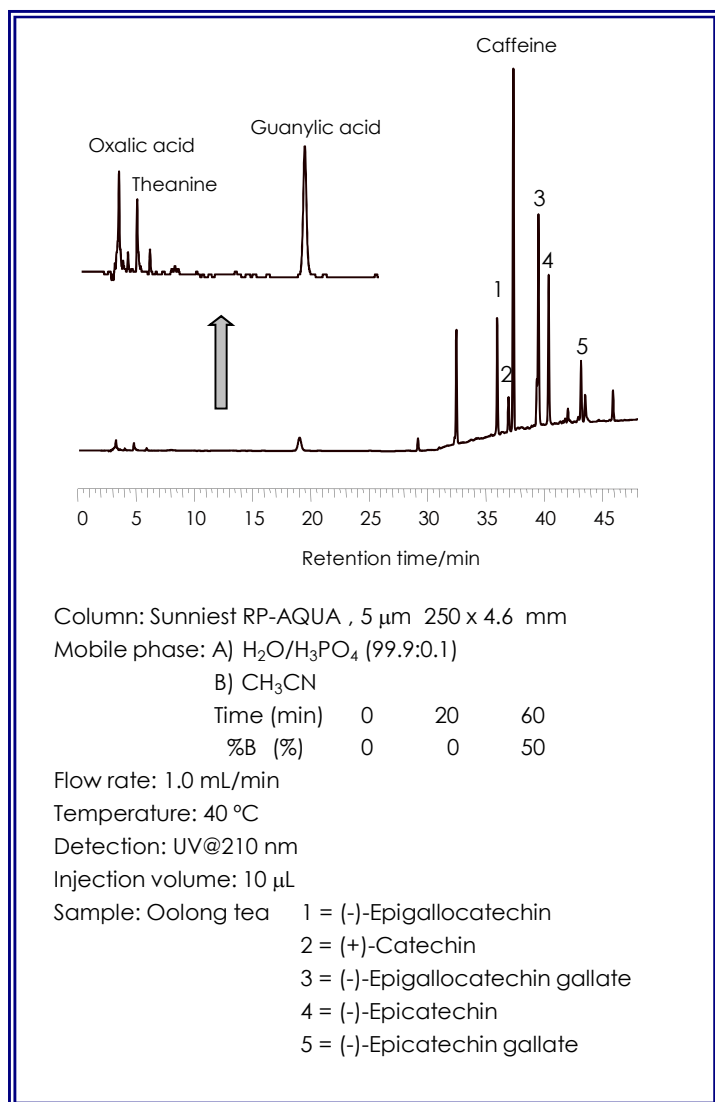
◆ Separation of water-soluble vitamins



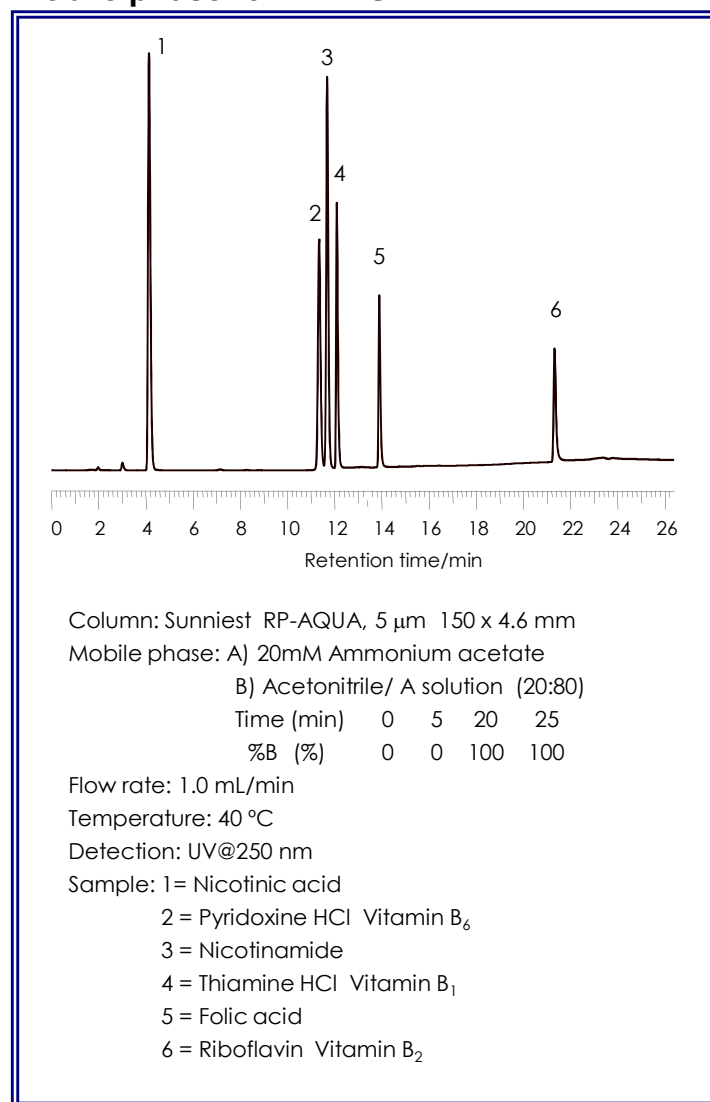
◆ Separation of nucleotides



◆ separation of Oolong tea



◆ Separation of water-soluble vitamins using mobile phase for LC/MS

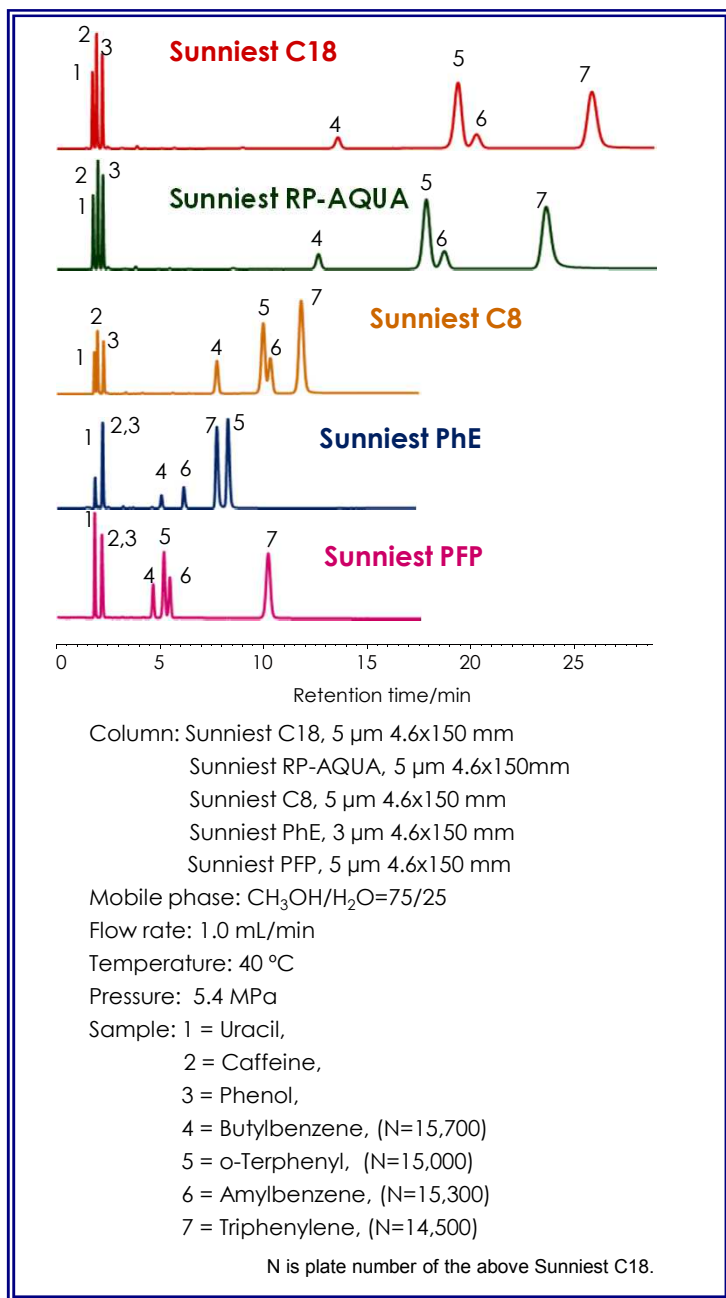


Sunniest C18

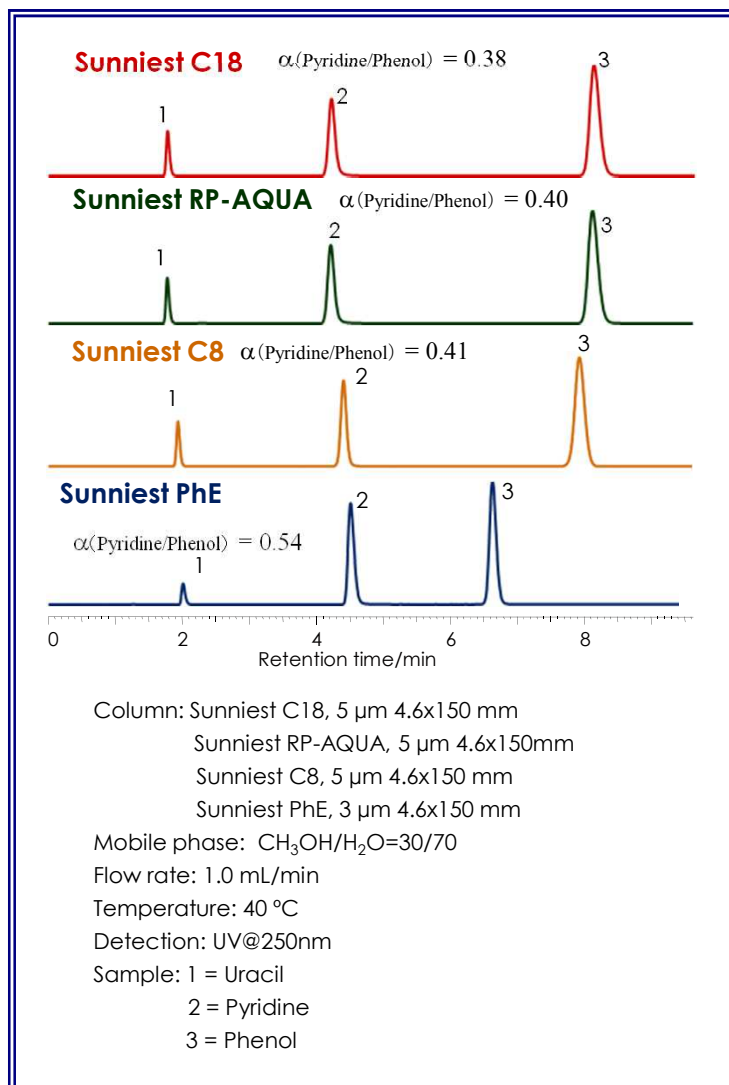
Sunniest RP-AQUA

Sunniest C8

◆ Separation of standard samples



◆ Separation of pyridine and phenol



Separation factor of pyridine and phenol is said to show the amount of residual silanol groups. The lower a value of separation factor, the less an effect of residual silanol groups.

All Sunniest columns show one of the lowest value.

| | C18 | RP-AQUA | C8 | PhE | PFP |
|-------------------------------------|------|---------|------|------|------|
| Hydrophobicity: | | | | | |
| α (Amylbenzene/Butylbenzene) | 1.56 | 1.56 | 1.43 | 1.34 | 1.29 |
| Hydrogen bonding capacity: | | | | | |
| α (Caffeine/Phenol) | 0.43 | 0.49 | 0.33 | 1.00 | 1.00 |
| Steric selectivity: | | | | | |
| α (Triphenylene/o-Terphenyl) | 1.37 | 1.36 | 1.23 | 0.92 | 2.51 |

Sunniest C18 shows not only high efficiency but also low column pressure.

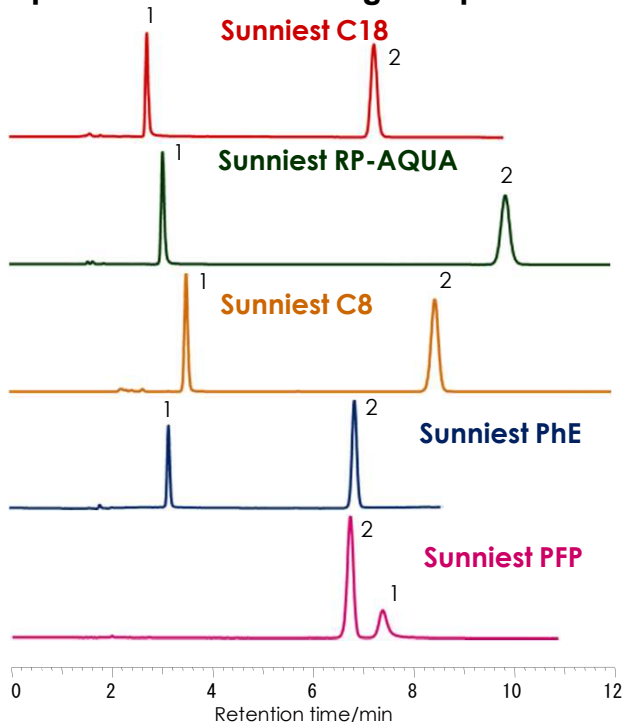


Sunniest C18

Sunniest RP-AQUA

Sunniest C8

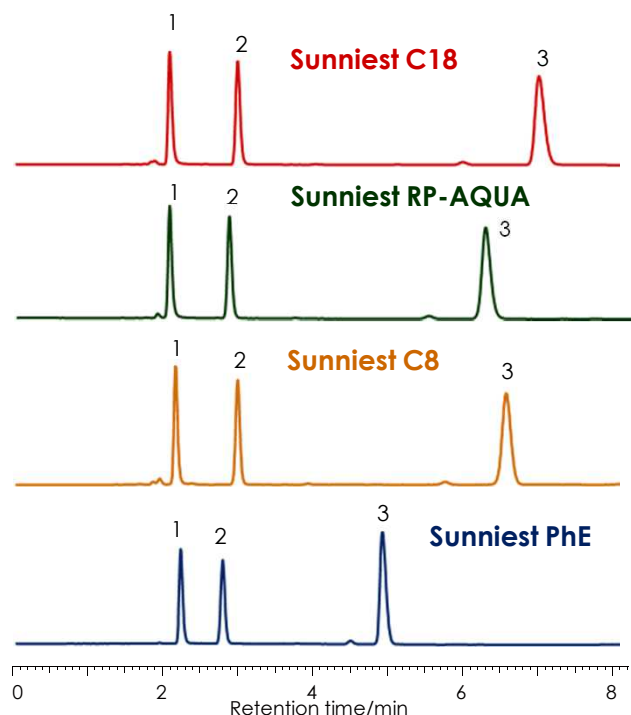
◆ Separation of a chelating compound



Column: Sunniest C18, 5 μ m 4.6x150 mm
 Sunniest RP-AQUA, 5 μ m 4.6x150mm
 Sunniest C8, 5 μ m 4.6x150 mm
 Sunniest PhE, 3 μ m 4.6x150 mm
 Sunniest PFP, 5 μ m 4.6x150 mm
 Mobile phase: CH₃CN/20mM H₃PO₄=10/90
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Detection: UV@250nm
 Sample: 1 = 8-Quinololinol
 2 = Caffeine

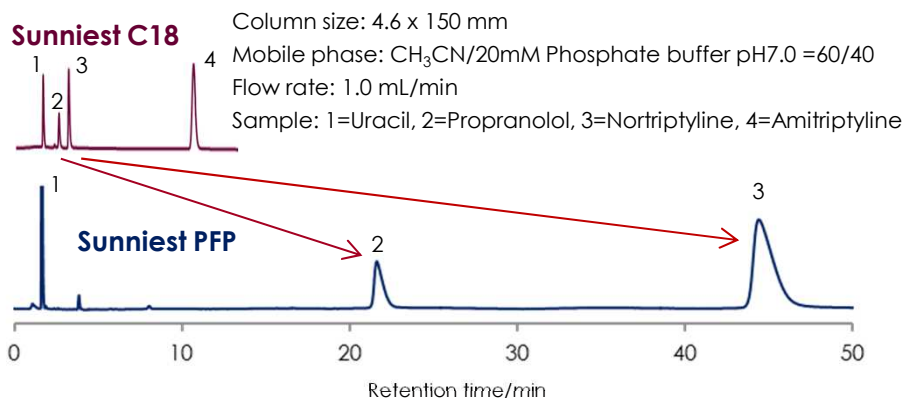
★ Sunniest C18, RP-AQUA, C8, PhE and PFP are inert for a metal chelating compound and acidic and basic compounds, so that they show symmetrical peaks of each compound.

◆ Separation of acidic compounds



Column: Sunniest C18, 5 μ m 4.6x150 mm
 Sunniest RP-AQUA, 5 μ m 4.6x150mm
 Sunniest C8, 5 μ m 4.6x150 mm
 Sunniest PhE, 3 μ m 4.6x150 mm
 Mobile phase: CH₃CN/0.1% H₃PO₄=2/98
 Flow rate: 1.0 mL/min
 Temperature: 40 °C
 Detection: UV@210nm
 Sample: 1 = Formic acid
 2 = Acetic acid
 3 = Propionic Acid

◆ Retention comparison between C18 and PFP



★ PFP retains a cation such a nortriptyline much longer than a C18.





Sunniest C18, C18-HT

Sunniest RP-AQUA

Sunniest C8

Sunniest PhE

Sunniest PFP

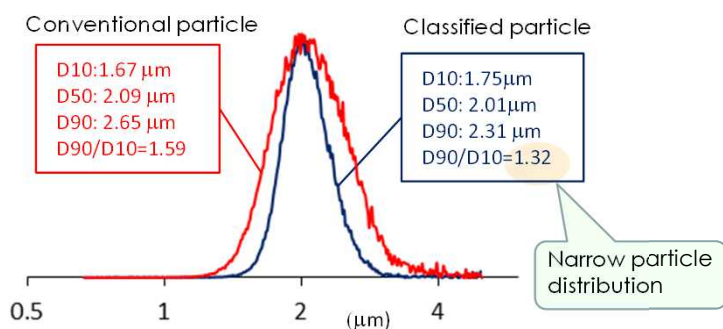
Sunniest C18-HT, 2 μm

Features

- Low back pressure and high efficiency by precisely classified particle
- High pressure packing (10,000 psi) using hard silica gels with high pressure resistant
- leads long column life without any void.
- Unique bonding technique for Sunniest
- The most suitable inner surface of column by special grinding

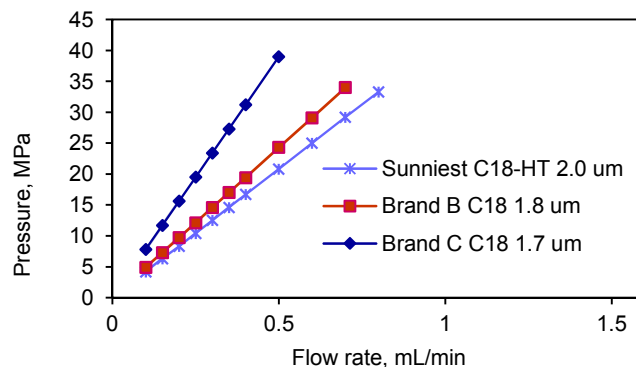
• Narrow Particle Distribution and Low Back Pressure

Measured by Coulter Counter method



Conventional 2 μm silica gel particle was classified again. 20% volume was cut off from both sides respectively. Consequently column back pressure reduced more than 15%. Our 2 μm silica gel particle shows a half pressure to compare with the other sub-2 μm silica gel particle.

Comparison of back pressure

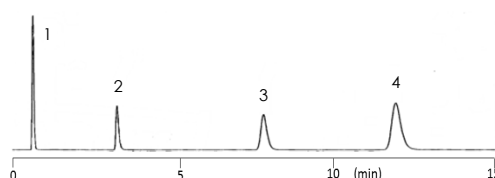


Column: Sunniest, Acquity and Zorbax
Column dimension: 50 x 2.1 mm
Mobile phase: Acetonitrile/water=(70/30)
Temperature: 25 $^{\circ}\text{C}$

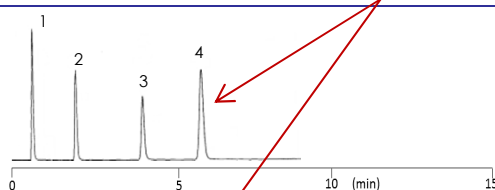
• An Unique Modification Leads Good Peak Shape

Neutral compounds

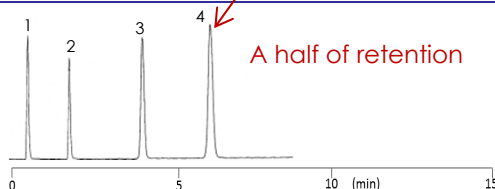
Sunniest C18-HT
2 μm



Acquity C18 1.7 μm

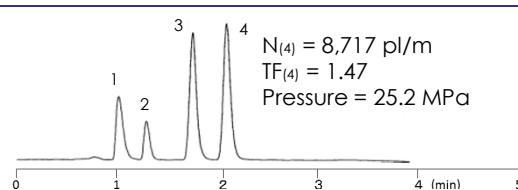


Leading C18
2.5 μm

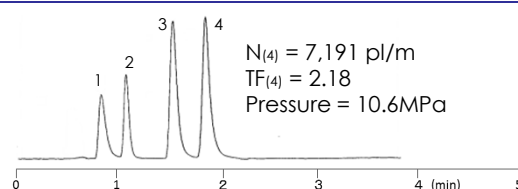


Amitriptyline (4th peak)

$N_{(4)} = 12,517 \text{ pl/m}$
 $TF_{(4)} = 1.29$
Pressure = 16.8 MPa

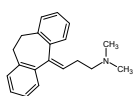


$N_{(4)} = 8,717 \text{ pl/m}$
 $TF_{(4)} = 1.47$
Pressure = 25.2 MPa



$N_{(4)} = 7,191 \text{ pl/m}$
 $TF_{(4)} = 2.18$
Pressure = 10.6 MPa

Column dimension: 50 x 2.1 mm
Mobile phase: Methanol/water=70/30 for neutral compounds
Methanol/25mM phosphate buffer (pH6.0)=80/20 for antidepressants
Flow rate: 0.2 mL/min
Temperature: room temperature
Sample: Neutral compounds, 1=Uracil, 2=Toluene, 3=Biphenyl, 4=Penanthrene
Antidepressants, 1=Nortriptyline, 2=Toluene, 3= Imipramine, 4=Amitriptyline



It is difficult to end-cap on sub 2 μm or 2 μm silica gel particle as well as 3 μm or 5 μm silica gel particle. Most sub 2 μm or 2 μm C18 columns show smaller plate number and larger tailing factor for a basic compound than Sunniest C18-HT. Sunniest C18-HT 2 μm shows a good peak shape for amitriptyline under methanol/phosphate buffer mobile phase at room temperature. Furthermore Sunniest C18-HT 2 μm shows 2 times longer retention time than the other brand columns.



Sunniest C18, C18-HT

Sunniest RP-AQUA

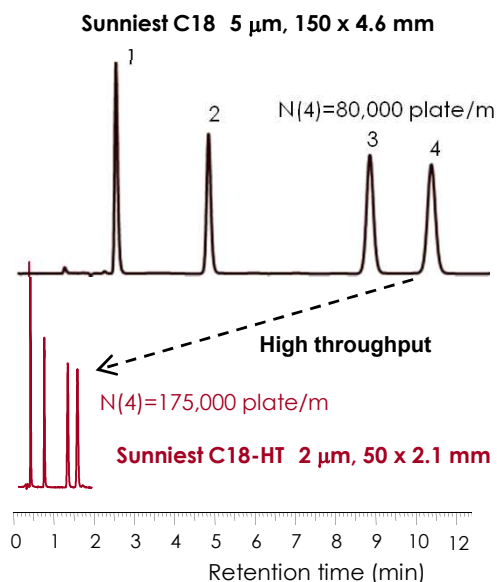
Sunniest C8

Sunniest PhE

Sunniest PFP

Sunniest C18-HT, 2 μ m

• Separation of Analgesics

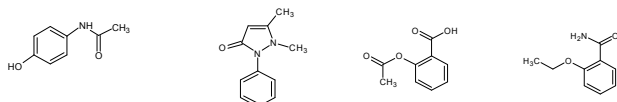


Mobile phase: CH₃CN/0.1% Formic acid = 20/80
Flow rate: 1.0 mL/min for 150 x 4.6 mm

0.6 mL/min for 50 x 2.1 mm

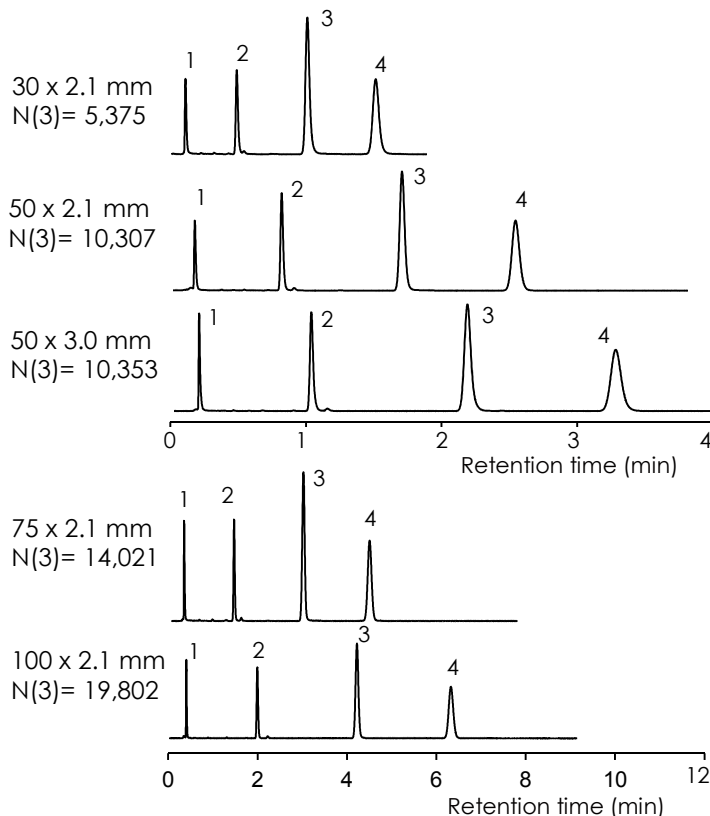
Temperature: 40 °C
Detection: UV@230 nm

Sample:
1=Acetaminophen, 2=Antipyrine, 3=Aspirin, 4=Ethenzamide



2 μ m particle allows to reduce retention time because high efficiency is kept under high flow rate conditions. As shown the above chromatograms, analytical time reduced 1/8 without sacrifices of separation by using 2 μ m, 50 x 2.1 mm column instead of 5 μ m 150 x 4.6 mm column.

• Comparison of Plate Number



Mobile phase: CH₃CN/H₂O = 60/40

Flow rate: 0.6 mL/min for 2.1 x 30 mm and 2.1 x 50 mm

1.0 mL/min for 3.0 x 50 mm

0.4 mL/min for 2.1 x 75 mm and 2.1 x 100 mm

Temperature: 40 °C

Detection: UV@250 nm

Sample: 1=Uracil,
2=Toluene,
3=Acenaphthene,
4=Butylbenzene

• Characteristics of Sunniest C18-HT, 2 μ m

| Packings | Silica gel support | | | C18 | | | |
|-----------------|--------------------------|--------------------|---|--------------------|--------------|----------------------------|--------------------|
| | Particle size (μ m) | Pore diameter (nm) | Specific surface area (m ² /g) | Carbon content (%) | Bonded phase | Maximum operating pressure | Available pH range |
| Sunniest C18-HT | 2.0 (Coulter counter) | 10 | 340 | 16 | C18 | 70 MPa or 10,000 psi | 1.5 - 10 |

It is very important for 2 μ m particle to have a capacity to resist pressure because of high column back pressure. The larger a pore volume of silica gel, the weaker a capacity to resist pressure. The silica gel with 0.85 ml/g of pore volume is used for Sunniest C18-HT, 2 μ m, so that it have a high capacity to resist pressure and a high operating pressure.



Sunniest C18, C18-HT

Sunniest RP-AQUA

Sunniest C8

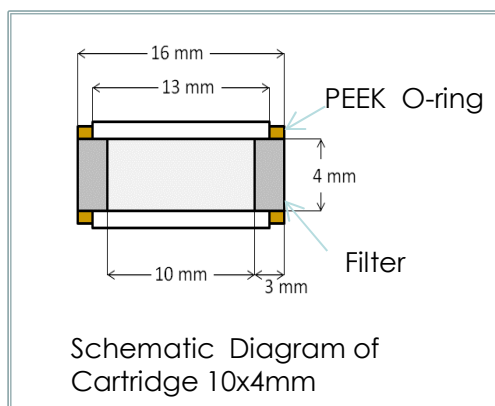
Sunniest PhE

Sunniest PFP

Guard Cartridge (10 x 4 mm)

Feature

- *Simple structure
- *Low dead volume
- *Available for not only 5 μ m column but also 3 μ m column



Comparison of chromatograms

Analytical column only

N(4) = 10,690
TF(4) = 1.05

Both Guard and Analytical columns

N(4) = 10,770
TF(4) = 1.07

Retention time/min

Column: Sunniest C18, 5 μ m 150 x 4.6 mm
Guard cartridge 10 x 4 mm

Mobile phase:

CH₃OH/20mM Phosphate buffer pH7.5 = 80/20

Flow rate: 1.0 mL/min

Temperature: 40 °C

Pressure: 4.8 MPa, 5.6 MPa(+guard)

Sample: 1 = Uracil,

2 = Propranolol,

3 = Nortriptyline,

4 = Amitriptyline,

TF: USP tailing factor



| | Particle size | Catalog No. |
|---|---------------|-------------|
| Sunniest C18, 5 μ m Guard cartridge column (1-pak + Holder) 4 x 10 mm | 5 μ m | EB3A1H |
| Sunniest RP-AQUA, 5 μ m Guard cartridge column (1-pak + Holder) 4 x 10 mm | 5 μ m | ER3A1H |
| Sunniest C8, 5 μ m Guard cartridge column (1-pak + Holder) 4 x 10 mm | 5 μ m | EC3A1H |
| Sunniest C18, 5 μ m Guard cartridge (4-pak) 4 x 10 mm | 5 μ m | EB3A1C |
| Sunniest RP-AQUA, 5 μ m Guard cartridge (4-pak) 4 x 10 mm | 5 μ m | ER3A1C |
| Sunniest C8, 5 μ m Guard cartridge (4-pak) 4 x 10 mm | 5 μ m | EC3A1C |
| Sunniest Guard cartridge holder 4 x 10 mm | --- | HOLA1C |

*** Sunniest Ordering information**

| Inner diameter [mm] | Length [mm] | Sunniest C18, 3 μ m | Sunniest C18, 5 μ m | Sunniest RP- AQUA, 3 μ m | Sunniest RP- AQUA, 5 μ m | Sunniest C8, 3 μ m | Sunniest C8, 5 μ m |
|------------------------|----------------|----------------------------|----------------------------|---------------------------------|---------------------------------|---------------------------|---------------------------|
| | | Catalog No. | Catalog No. | Catalog No. | Catalog No. | Catalog No. | Catalog No. |
| 2 | 50 | EB2241 | EB3241 | ER2241 | ER3241 | EC2241 | EC3241 |
| | 75 | EB2251 | — | ER2251 | — | EC2251 | — |
| | 100 | EB2261 | EB3261 | ER2261 | ER3261 | EC2261 | EC3261 |
| | 150 | EB2271 | EB3271 | ER2271 | ER3271 | EC2271 | EC3271 |
| | 250 | EB2281 | EB3281 | ER2281 | ER3281 | EC2281 | EC3281 |
| 3 | 50 | EB2341 | EB3341 | ER2341 | ER3341 | EC2341 | EC3341 |
| | 100 | EB2361 | EB3361 | ER2361 | ER3361 | EC2361 | EC3361 |
| | 150 | EB2371 | EB3371 | ER2371 | ER3371 | EC2371 | EC3371 |
| | 250 | EB2381 | EB3381 | ER2381 | ER3381 | EC2381 | EC3381 |
| 4.6 | 10 | EB2411 | EB3411 | ER2411 | ER3411 | EC2411 | EC3411 |
| | 50 | EB2441 | EB3441 | ER2441 | ER3441 | EC2441 | EC3441 |
| | 75 | EB2451 | — | ER2451 | — | EC2451 | — |
| | 100 | EB2461 | EB3461 | ER2461 | ER3461 | EC2461 | EC3461 |
| | 150 | EB2471 | EB3471 | ER2471 | ER3471 | EC2471 | EC3471 |
| | 250 | EB2481 | EB3481 | ER2481 | ER3481 | EC2481 | EC3481 |
| 10 | 250 | — | EB3781 | — | ER3781 | — | EC3781 |
| 20 | 50 | — | EB3841 | — | ER3841 | — | EC3841 |
| | 150 | — | EB3871 | — | ER3871 | — | EC3871 |
| | 250 | — | EB3881 | — | ER3881 | — | EC3881 |

| Inner diameter [mm] | Length [mm] | Sunniest PhE, 3 μ m | Sunniest PhE, 5 μ m | Sunniest PFP, 5 μ m |
|------------------------|----------------|----------------------------|----------------------------|----------------------------|
| | | Catalog No. | Catalog No. | Catalog No. |
| 2.0 | 50 | EP2241 | EP3241 | — |
| | 75 | EP2251 | — | — |
| | 100 | EP2261 | EP3261 | — |
| | 150 | EP2271 | EP3271 | — |
| | 250 | EP2281 | EP3281 | — |
| 3.0 | 50 | EP2341 | EP3341 | — |
| | 100 | EP2361 | EP3361 | — |
| | 150 | EP2371 | EP3371 | — |
| | 250 | EP2381 | EP3381 | — |
| 4.6 | 10 | — | EP3411 | — |
| | 50 | EP2441 | EP3441 | EF3441 |
| | 75 | EP2451 | — | — |
| | 100 | EP2461 | EP3461 | EF3461 |
| | 150 | EP2471 | EP3471 | EF3471 |
| | 250 | EP2481 | EP3481 | EF3481 |
| 10.0 | 250 | — | EP3781 | — |
| 20.0 | 50 | — | EP3841 | — |
| | 150 | — | EP3871 | — |
| | 250 | — | EP3881 | — |

| Inner diameter [mm] | Length [mm] | Sunniest C18-HT, 2 μ m |
|------------------------|----------------|-------------------------------|
| | | Catalog No. |
| 2.1 | 20 | EB1921 |
| | 30 | EB1931 |
| | 50 | EB1941 |
| | 75 | EB1951 |
| | 100 | EB1961 |
| 3.0 | 20 | EB1321 |
| | 30 | EB1331 |
| | 50 | EB1341 |
| | 75 | EB1351 |
| | 100 | EB1361 |